

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/230621744>

# Aspirin Inhibits Oxidant Stress, Reduces Age-Associated Functional Declines, and Extends Lifespan of *Caenorhabditis elegans*

Article in *Antioxidants and Redox Signaling* · August 2012

DOI: 10.1089/ars.2011.4151 · Source: PubMed

CITATIONS

83

READS

492

8 authors, including:



**Srinivas Ayyadevara**

University of Arkansas for Medical Sciences

88 PUBLICATIONS 1,885 CITATIONS

[SEE PROFILE](#)



**Abhijit Dandapat**

31 PUBLICATIONS 1,696 CITATIONS

[SEE PROFILE](#)



**Chang-Ping Hu**

Central South University

141 PUBLICATIONS 4,348 CITATIONS

[SEE PROFILE](#)



**Magomed Khaidakov**

University of Arkansas for Medical Sciences

82 PUBLICATIONS 1,836 CITATIONS

[SEE PROFILE](#)

Some of the authors of this publication are also working on these related projects:



Thiamine Therapy for Heart Failure: a Promise or Fiction? [View project](#)



Protein aggregation and its roles in neurodegeneration and other age-progressive diseases. [View project](#)

ORIGINAL RESEARCH COMMUNICATION

# Aspirin Inhibits Oxidant Stress, Reduces Age-Associated Functional Declines, and Extends Lifespan of *Caenorhabditis elegans*

Srinivas Ayyadevara,<sup>1,2,\*</sup> Puneet Bharill,<sup>3,\*</sup> Abhijit Dandapat,<sup>1,4,\*</sup> Changping Hu,<sup>1,4</sup> Magomed Khaidakov,<sup>4</sup> Sona Mitra,<sup>4</sup> Robert J. Shmookler Reis,<sup>1-3</sup> and Jawahar L. Mehta<sup>1,4</sup>

## Abstract

**Aims:** Oxidative stress and inflammation are leading risk factors for age-associated functional declines. We assessed aspirin effects on endogenous oxidative-stress levels, lifespan, and age-related functional declines, in the nematode *Caenorhabditis elegans*. **Results:** Both aspirin and its salicylate moiety, at nontoxic concentrations (0.5–1 mM), attenuated endogenous levels of reactive oxygen species ( $p < 0.001$ ), and upregulated antioxidant genes encoding superoxide dismutases (especially *sod-3*,  $p < 0.001$ ), catalases (especially *ctl-2*,  $p < 0.0001$ ), and two glutathione-S-transferases (*gst-4* and *gst-10*; each  $p < 0.005$ ). Aspirin, and to a lesser degree salicylate, improved survival of hydrogen peroxide, and in the absence of exogenous stress aspirin extended lifespan by 21%–23% (each  $p < 10^{-9}$ ), while salicylate added 14% ( $p < 10^{-6}$ ). Aspirin and salicylate delayed age-dependent declines in motility and pharyngeal pumping (each  $p < 0.005$ ), and decreased intracellular protein aggregation ( $p < 0.0001$ )—all established markers of physiological aging—consistent with slowing of the aging process. Aspirin fails to improve stress resistance or lifespan in nematodes lacking DAF-16, implying that it acts through this FOXO transcription factor. **Innovation:** Studies in mice and humans suggest that aspirin may protect against multiple age-associated diseases by reducing all-cause mortality. We now demonstrate that aspirin markedly slows many measures of aging in the nematode. **Conclusions:** Aspirin treatment is associated with diminished endogenous oxidant stress and enhanced resistance to exogenous peroxide, both likely mediated by activation of antioxidant defenses. Our evidence indicates that aspirin attenuates insulin-like signaling, thus protecting against oxidative stress, postponing age-associated functional declines and extending *C. elegans* lifespan under benign conditions. *Antioxid. Redox Signal.* 18, 481–490.

## Introduction

AGING IS A multi-factorial process influenced by environmental and genetic factors. Many studies have indicated that oxidative damage limits longevity (15, 31, 50), although some have questioned a causal role of oxidation in aging due to failure of antioxidant interventions to extend life (12, 36, 46). Nonetheless, there is strong evidence that aging-related declines in mitochondrial function and integrity are associated with oxidative stress (5), and the role of oxidative damage in aging itself remains controversial (50).

A complex interplay of reactive oxygen species (ROS) generation and genes that protect against ROS damage has been implicated in the onset and progression of numerous

aging-associated disease states, including type-2 diabetes, solid cancers, Alzheimer's dementia, osteoporosis, myocardial infarction, and cerebrovascular accidents (45). Blockade of insulin/IGF-1 signaling has beneficial effects: it increases lifespan, confers resistance to oxidative stress, and reduces or postpones many age-associated conditions and diseases (2, 8, 17, 38). How could different stress states, including those mediated by ROS, oxidants and other electrophiles, contribute to aging and explain the age-dependent incidence of diverse diseases? A possible shared mechanism, increasingly invoked in both geriatric and gerontologic contexts, is inflammation (13).

Aspirin (acetylsalicylic acid), a prototypic cyclooxygenase inhibitor, is a widely used analgesic agent which opposes

<sup>1</sup>Central Arkansas Veterans Healthcare System, Little Rock, Arkansas.

Departments of <sup>2</sup>Geriatrics, <sup>3</sup>Biochemistry and Molecular Biology, and <sup>4</sup>Medicine, University of Arkansas for Medical Sciences, Little Rock, Arkansas.

\*These authors contributed equally to this work.

### Innovation

Aspirin is a versatile drug that confers protection against multiple age-associated diseases, including atherosclerosis, diabetes, and a variety of cancers. Evidence in mice and humans suggests that aspirin modestly reduces all-cause mortality, but it is unknown whether that reduction is merely the sum of the disease-specific benefits or a more direct amelioration of aging itself. Our observations demonstrate for the first time that aspirin slows many measures of aging *per se*, and provides evidence that insulin-like signaling and antioxidant defenses are involved. The nematode model is a particularly suitable one in which to establish the mechanisms underlying aspirin's diverse benefits.

inflammation and platelet aggregation. It regulates the activity of a number of pro-inflammatory signaling molecules, such as TGF- $\beta$  (37) and PDGF (48). In addition to its anti-inflammatory effect mediated through cyclooxygenase inhibition, the salicylate moiety in aspirin also reduces the generation of pro-oxidant species (28). In keeping with these pharmacologic effects, aspirin reduces the severity of aging-related endothelial dysfunction in mice (7), modestly lowers blood pressure in hypertensive mice (26), and reduces cardiovascular events in both primary and secondary prevention trials in humans (11). High-dose aspirin was reported to improve glucose metabolism and reduce fatty acid levels in patients with type-2 diabetes (20). Aspirin also extends the lifespan of male mice (43), and significantly reduces all-cause mortality in patients with type-2 diabetes (35). The mechanisms by which aspirin increases longevity, however, remain unclear.

The present study was designed to assess the effects of aspirin on endogenous ROS levels and regulation of antioxidant genes in a simple model system, *Caenorhabditis elegans*. Superoxide dismutases (SODs) convert superoxide into oxygen and peroxide, which in turn is broken down by catalases to molecular oxygen and water. Peroxide can trigger quite harmful free-radical chain reactions in cell lipids, and thus is likely to be the main source of oxidative damage associated with aging (41), opposed chiefly by certain glutathione S-transferases (GSTs) (50). We now demonstrate protective effects of aspirin, accompanied by induction of antioxidant enzymes (SODs, catalases, and lipoperoxidation-specific GSTs). Moreover, we document its ability to attenuate aging-associated functional declines and to extend lifespan in the nematode, largely dependent on the FOXO transcription factor that mediates most downstream effects of insulin-like signaling.

### Results

#### *Aspirin and salicylate reduce endogenous oxidative stress, and upregulate antioxidant enzymes and phase-2 detoxification enzymes*

*In vivo* steady-state ROS levels can be inferred from fluorogenic activation of dichlorodihydrofluorescein diacetate (H<sub>2</sub>DCF-DA, Invitrogen/Life Technologies), provided that the intensity and duration of excitation (incident light) are held rigorously constant (10). Total H<sub>2</sub>DCF-DA fluorescence was reduced ~40% by treating worms with either 1 mM aspirin or 1 mM salicylate (each  $p < 0.001$  vs. untreated controls; Fig. 1A and 1B). H<sub>2</sub>DCF-DA responds most sensitively to hydroxyl radical (www.invitrogen.com).

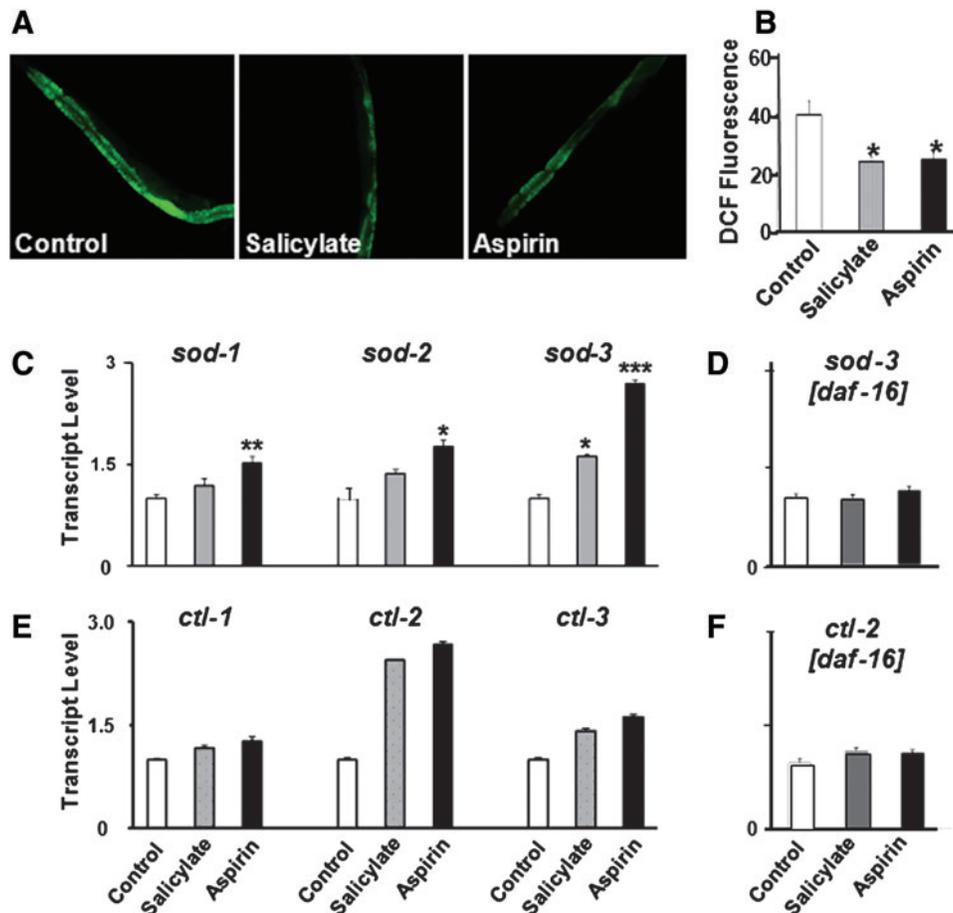
Because neither aspirin nor salicylate is itself an antioxidant, we sought mechanisms by which they might reduce ROS abundance. Transcript levels were measured for three distinct classes of antioxidant enzymes: SODs, catalases, and GSTs. With one exception, all were induced by aspirin and to a lesser extent by salicylate, each at 1 mM concentration. Of the genes encoding SODs (Fig. 1C), the most strongly induced (1.6-fold by salicylate; 2.7-fold by aspirin,  $p < 0.0003$ ) was *sod-3*, encoding an Fe/Mn-SOD believed to be localized to mitochondria. *Sod-4*, encoding an extracellular Cu/Zn-SOD, was downregulated 3.5-fold by 1 mM aspirin ( $p < 10^{-4}$ ), while *sod-5* transcripts were increased ~20% (Supplementary Fig. S1; supplementary data are available online at www.liebertonline.com/ars). The most affected catalase gene (Fig. 1E) was *ctl-2*, induced 2.4-fold by salicylate ( $p < 0.001$ ) and 2.7-fold by aspirin ( $p < 10^{-4}$ ). CTL-2 protein is found primarily in intestinal peroxisomes. Of the five GST genes tested (data not shown), the most affected were *gst-10*, upregulated 2.5-fold by aspirin ( $p < 0.001$ ) and *gst-5*, induced ~2-fold by aspirin ( $p < 0.005$ ). No upregulation of *sod* or *ctl* genes was observed in a *daf-16* mutant (e.g., see Fig. 1D and 1F), implying that these effects of aspirin treatment require the DAF-16/FOXO transcription factor.

Consistent with transcriptional regulation via DAF-16, the most affected antioxidant genes are believed to be negatively regulated by insulin/IGF-1 signaling, and show increased expression in long-lived *daf-2* mutants in which the insulin-like receptor is disrupted (18). The most affected *gst* genes, *gst-5* and *gst-10*, were the only two of 44 genes tested that extend lifespan when disrupted (3). We examined the expression of GFP-reporter constructs driven by promoters for *gst-4* and *gst-10*, encoding GSTs that protect against lipoperoxidation; their expression is also upregulated in *daf-2* mutants (27), although chiefly via the SKN-1 transcription factor that is closely intertwined with insulin-like signaling (34). Expression of P<sub>*gst-4*</sub>::GFP and P<sub>*gst-10*</sub>::GFP increased at least 3.5-fold in aspirin-treated worms relative to untreated controls (Fig. 2; each  $p < 0.0001$ ), corroborating aspirin induction of antioxidant defenses. Expression of the P<sub>*sod-3*</sub>::GFP reporter in transgenic worms was increased at least 1.6-fold by aspirin (Fig. 2, bottom panels), somewhat less than the upregulation observed at the transcript level by RT-PCR (Fig. 1C).

#### *Aspirin improves resistance to an oxidative stress and extends C. elegans lifespan*

Wild-type *C. elegans* were exposed to aspirin or salicylate from the last larval stage (L4, 2 days after hatching from eggs), and transferred on day 5 post-hatch to liquid medium containing a toxic level (5 mM) of H<sub>2</sub>O<sub>2</sub>. As shown in Figure 3A, aspirin and to a lesser extent salicylate (each at 1 mM) significantly extended survival of wild-type (N2) nematodes in the presence of a toxic level of H<sub>2</sub>O<sub>2</sub> ( $p < 0.001$  and  $p < 0.01$ , respectively).

Because stress resistance can provide a surrogate biomarker for longevity (22, 29, 40–42), we asked whether aspirin might also extend nematode lifespan. Aspirin (0.5 or 1.0 mM) increased the lifespan of wild-type worms by 12%–30% in three experiments (each  $p < 0.001$ , data summarized in Table 1). A typical set of survivals is shown in Figure 3C; overall, the weighted average life extensions by 0.5 and 1.0 mM aspirin were 23% and 21%, respectively, with a combined  $p$  value of  $< 10^{-9}$  at 1 mM (Table 1). The salicylate moiety of aspirin, at 1 mM, also enhanced *C. elegans* lifespan in three experiments (e.g., Fig. 3C), producing an average extension of 14% (combined  $p < 10^{-7}$ ; Table 1).



**FIG. 1. Reduction of oxidative stress by aspirin and salicylate.** (A, B) Steady-state levels of reactive oxygen species (ROS), as reflected by DCF fluorescence, were significantly reduced by aspirin or salicylate treatment (each at 1 mM). (A) shows typical worms in each group, while (B) summarizes data for total fluorescence per worm ( $\geq 20$  worms per group). \*Differs from control at  $p < 0.001$  by 2-tailed  $t$ -test. (C–F) Steady-state transcript levels of catalase (*ctl*) and superoxide dismutase (*sod*) oxidative-defense genes, in the presence of 1 mM aspirin, 1 mM salicylate, or vehicle. Transcript levels were assessed by RT-PCR, relative to  $\beta$ -actin transcript abundance in the same group. Bars indicate means  $\pm$  SEM for 3–5 independent biological expansions per group. (C) Three of the nematode superoxide dismutase (*sod*) genes in wild-type worms, strain Bristol N2-DRM; (D) *sod-3* transcripts in *daf-16(mu86)* worms; (E) the three nematode catalase (*ctl*) genes in wild-type worms, strain Bristol-N2/DRM; (F) *ctl-2* transcripts in *daf-16(mu86)* worms. Significance of differences from control, by 2-tailed  $t$ -test for small samples or samples of unequal variance (appropriate for small  $N$ ):  $^{\ddagger}p < 0.05$ ;  $^*p < 0.02$ ;  $^{**}p < 0.01$ ;  $^{***}p < 0.001$ ;  $^{****}p < 0.0001$ . (To see this illustration in color, the reader is referred to the web version of this article at [www.liebertpub.com/ars](http://www.liebertpub.com/ars).)

Insulin/IGF-1 signaling has been implicated in aging of *C. elegans*, results replicated in *Drosophila* and mice (14), and is almost entirely dependent on the DAF-16/FOXO transcription factor (25). We assessed whether lifespan extension and stress resistance conferred by aspirin also require DAF-16. As shown in Figure 3B and 3D (data summarized in Table 1), aspirin had no detectable effect on either survival of  $H_2O_2$  stress, or lifespan, for a *C. elegans* mutant in which the *daf-16* gene was severely disrupted, implying that these salutary effects of aspirin depend, largely or entirely, on the DAF-16/FOXO transcription factor.

#### Aspirin delays age-associated functional declines in *C. elegans*

Aging is associated with a progressive decline in many physiological functions, such as spontaneous movement and

pharyngeal pumping in the nematode (16, 19). As shown in Figure 4A, the age-dependent decline in pharyngeal pumping was reduced or delayed by 1 mM aspirin or salicylate (each  $p < 0.001$  on day 6, and  $p < 0.05$  on day 10 of adulthood). In this respect, aspirin- and salicylate-treated worms appeared physiologically “younger” than the untreated controls. Since food intake was initially unchanged (at the first assay, on adult day 2), and was subsequently *higher* in the treated worms than in controls, it can be surmised that aspirin and salicylate do not exert their salutary effects via dietary restriction (*i.e.*, by suppressing food intake).

A progressive decline with age in *C. elegans* motility, both spontaneous and in response to exogenous stimuli, has been well documented (16, 19). Continuous exposure to either aspirin or salicylate (each at 1 mM) significantly inhibited the age-related decline in motility. The fraction of untreated worms moving in response to touch (Fig. 4B) fell 30% between

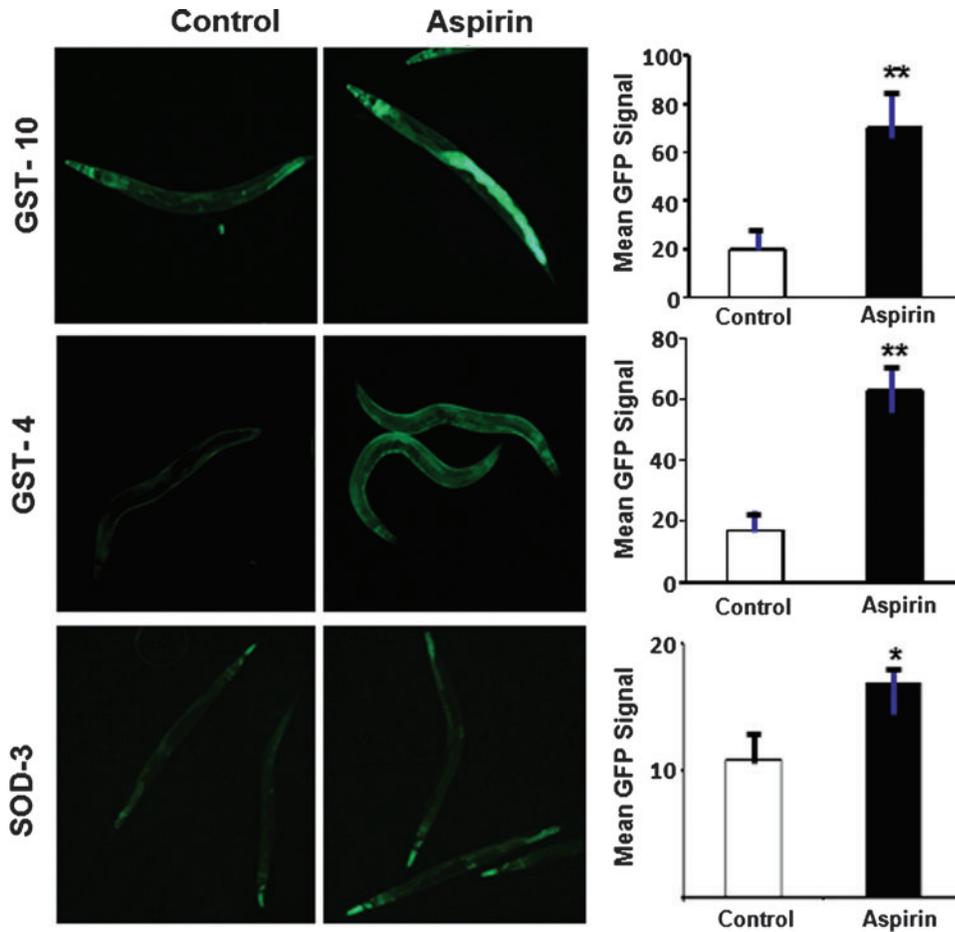


FIG. 2. Reporter strains globally expressing GFP driven by promoters from *gst-10*, *gst-4*, or *sod-3* genes. Typical examples are shown of fluorescence in adult wild-type worms. Histogram panels at right summarize quantitative whole-body fluorescence data for  $\geq 20$  worms per group. Significance of aspirin – control differences by 2-tailed *t*-test ( $N=20$ ): \* $p < 0.001$ ; \*\* $p < 0.0001$ . (To see this illustration in color, the reader is referred to the web version of this article at [www.liebertpub.com/ars](http://www.liebertpub.com/ars).)

10 and 14 days of adult age (Chi-squared  $p < 10^{-6}$ ); in contrast, the decline was  $< 3\%$  in aspirin- or salicylate-treated worms, not significantly changed from day 10 (open bars in Fig. 4B). At 14 days, however, ASA- and salicylate-treated worms showed significantly more motility than untreated controls (solid bars;  $P < 10^{-6}$  and 0.0005, respectively). Spontaneous movement was relatively infrequent at adult day 10 ( $\sim 8.6\%$ ), and declined to 0.3% by day 14 (Fig. 4C, Control bars). Treatment with aspirin or salicylate doubled the spontaneous movement frequency at the earlier time point (Fig. 4C, open bars; each  $p < 0.005$ ), and raised the 14-day value (filled bars) from 0.3% to 3.7% ( $p < 0.005$ ) and 2.3% ( $p < 0.05$ ), respectively. Both the pumping rate and motility data suggest that aspirin and salicylate slow the decline in function that accompanies physiological aging.

#### Aspirin reduces aging-associated aggregation of proteins in *C. elegans*

Aggregation of proteins, a characteristic of many progressive neuropathies, increases with age but is slowed by mutations that extend lifespan, and may thus provide a robust molecular biomarker of aging (6, 29). We assessed a transgenic strain expressing a fluorescence-tagged polyglutamine repeat ( $Q_{40}::YFP$ ) in body wall muscle (6). Such strains undergo age-dependent aggregate formation, as evidenced by a highly punctate pattern of yellow fluorescence in body wall muscle, commencing at day 3 after hatch (adult day 1). Aspirin reduced both the size and number of  $Q_{40}::YFP$  aggre-

gates observed at adult day 4, each by  $> 30\%$  ( $p < 0.0001$ ) (Fig. 5).

#### Discussion

Oxidative stress generally elicits an inflammatory response that, under favorable conditions, initiates tissue repair and restoration of a normal physiological state. However, in senescence, excessive oxidative stress is not adequately controlled by endogenous antioxidant reserves and may produce a state of chronic inflammation (13, 47). Aspirin, a very widely used drug, is a potent anti-inflammatory agent. We have shown that this drug, and to a somewhat lesser extent its salicylate moiety, can reduce age-associated oxidative stress and attenuate insulin/IGF-1 signaling (based on known, antioxidant gene targets of this pathway) in *C. elegans*. Both these agents also delay several measures of physiological aging and extend nematode lifespan.

One shared precept of various “oxidative stress theories of aging” is that endogenous ROS species, including superoxide and peroxide radical, either increase with age or are less effectively offset by antioxidant defenses, resulting in many aging-associated declines in tissue functions (42). In the present studies, we noted that both aspirin and salicylate attenuate, by  $> 40\%$ , total signal reported by the fluorogenic dye  $H_2DCF\text{-DA}$  as a measure of endogenous ROS levels (10). To evaluate the balance between ROS generation and subsequent elimination by antioxidant enzymes, we also measured the transcript levels of key “longevity-assurance genes”

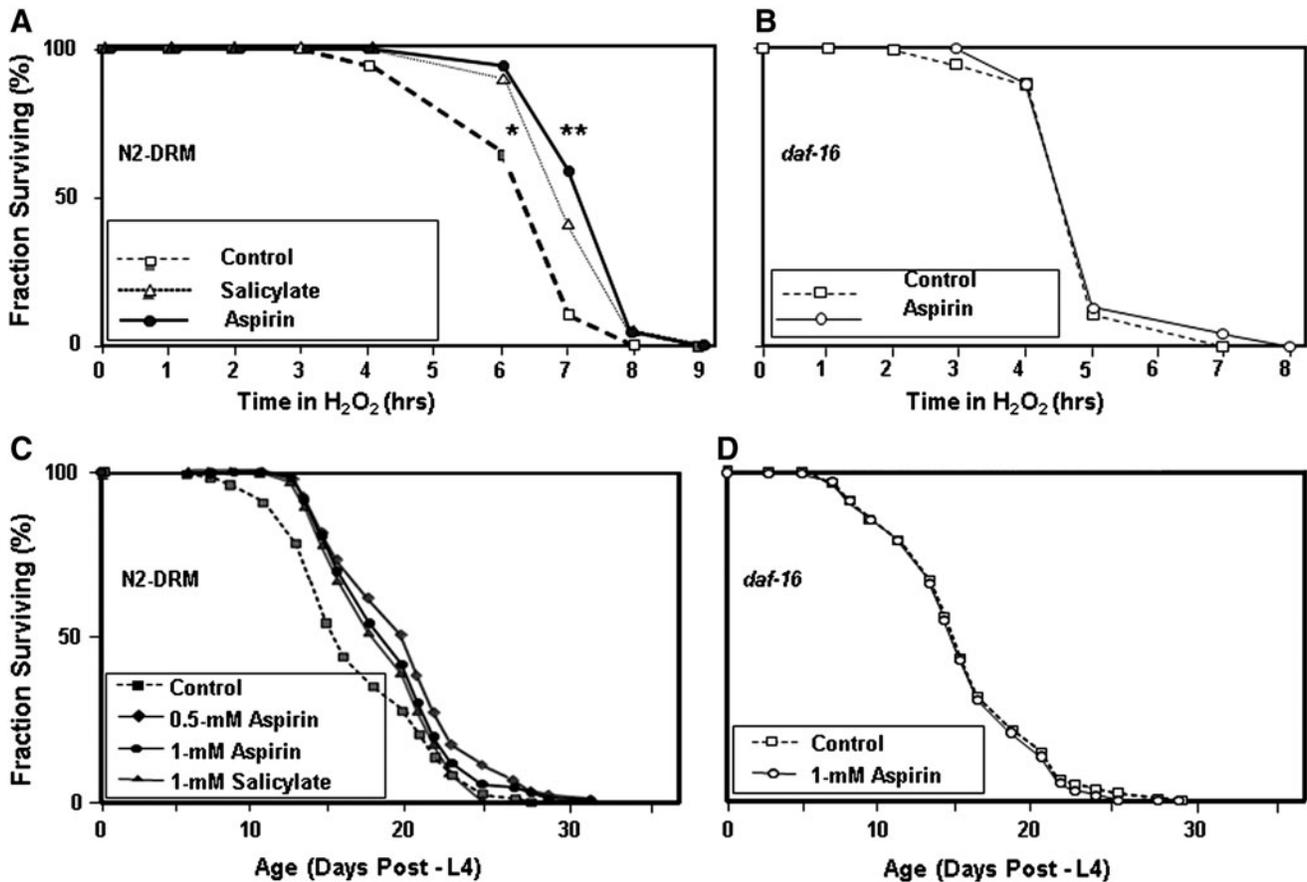


FIG. 3. Aspirin and salicylate extend *C. elegans* survival of oxidative stress and lifespan under benign conditions. Aspirin and salicylate (each at 1 mM) extended survival in 5 mM hydrogen peroxide for wild-type worms [N2-DRM, (A)], but had no effect on the *daf-16* (*mu86*) mutant strain (B). Treated N2 worms differed from control survivals by Gehans-Wilcoxon log-rank test: \* $p < 0.01$ ; \*\* $p \leq 0.001$ . Life-span of N2 wild-type worms was also extended by 0.5- or 1 mM aspirin or 1 mM salicylate [(C) each  $p < 0.001$  vs. control, by Gehans-Wilcoxon log-rank test;  $N \geq 90$  for each group], but the *daf-16* mutant derived no benefit from either drug (D). Replicate experiments all demonstrated significant extension of wild-type lifespan by aspirin and salicylate (3 independent tests) as summarized in Table 1; the experiment shown in (C) and (D) is "Experiment 1" of Table 1.

encoding 3 catalases, 5 SODs, and 5 GSTs. We found that aspirin, and to a lesser extent salicylate, increased transcript levels for nearly all of the tested antioxidant genes, with the greatest effect (>2.5-fold for aspirin) on *sod-3* and *ctl-2*. In keeping with these observations, we found that both aspirin and salicylate improved survival in the presence of exogenous H<sub>2</sub>O<sub>2</sub> (Fig. 3A). A curious exception to this pattern is presented by *sod-4*, which was downregulated more than 3-fold by aspirin (Supplementary Fig. S1). It was previously reported that deletion of the *sod-4* gene, which encodes an extracellular SOD and contributes ~5% of total *sod* transcripts (46), has no effect on lifespan or resistance to several oxidative stresses in a wild-type background, but further increases lifespan (especially maximal lifespan) in the long-lived *daf-2* background (12). Since aspirin appears to reduce insulin-like signaling, and thus resembles a *daf-2* hypomorphic mutation, it is likely that aspirin-mediated suppression of *sod-4* would serve to promote rather than impair longevity.

Peroxide is a potent inducer of lipid peroxidation, which poses the greatest oxidative threat to survival because lipoperoxides (unlike other ROS molecules) are sufficiently long-lived to create free-radical chain reactions *in vivo* (50). The two nematode glutathionyl S-transferases with highest activity

against lipoperoxides are GST-4 and GST-10 (3). It is thus particularly significant that protein-level expression of reporters for these GST's was increased >3-fold by aspirin treatment (Fig. 2). Considering that vulnerability to lipid peroxidation is one of the strongest predictors of nematode longevity (41), aspirin's ability to strongly induce these enzymes may contribute to its extension of nematode lifespan.

The GSTs are phase-2 detoxification genes, which have been proposed to play key roles in longevity assurance (21, 27). In addition to GSTs, other phase-1 and phase-2 enzymes mediating resistance to endogenous and exogenous toxicants (which include but extend beyond ROS) comprise the catalases, SODs, cytochromes P450, short-chain dehydrogenases/reductases, and UDP-glucuronosyltransferases (UGTs) (33). These genes are broadly upregulated in long-lived *C. elegans* mutants including *daf-2*, *age-1* and *clk-1*, which in particular overexpress *sod-3* and *ctl-2* (32), the two antioxidant genes most highly induced by aspirin and salicylate treatments (Fig. 1C and 1E), and reported to be essential for *daf-16*-mediated innate resistance to bacterial infection (9). Taken together, the evidence that aspirin treatment of *daf-16* mutant worms enhanced neither peroxide resistance nor lifespan (Fig. 3B and 3D), and failed to induce *sod-3* and *ctl-2* (Fig. 1D and 1F),

TABLE 1. ASPIRIN AND SALICYLATE EXTEND ADULT LIFESPAN OF *C. ELEGANS* AT 20°C

Strain	Condition	Deaths (N)	Mean $\pm$ SD	% Increase	p value*
<i>Experiment 1</i>					
N2DRM	Control (vehicle)	95	15.7 $\pm$ 2.7	—	—
N2DRM	Aspirin (0.5 mM)	100	19.4 $\pm$ 2.3	24	< 0.001
N2DRM	Aspirin (1 mM)	90	18.8 $\pm$ 2.3	20	< 0.001
N2DRM	Salicylate (1 mM)	98	17.6 $\pm$ 2.7	12	< 0.001
daf-16	Control (vehicle)	35	14.5 $\pm$ 2.8	—	—
daf-16	Aspirin (1 mM)	34	14.5 $\pm$ 1.8	0	0.81
<i>Experiment 2</i>					
N2DRM	Control (vehicle)	32	14.8 $\pm$ 4.7	—	—
N2DRM	Aspirin (0.5 mM)	35	19.2 $\pm$ 4.2	30	< 0.001
N2DRM	Aspirin (1 mM)	35	18.6 $\pm$ 4.3	26	< 0.001
N2DRM	Salicylate (1 mM)	31	18.1 $\pm$ 4.2	22	< 0.01
daf-16	Control (vehicle)	35	13.5 $\pm$ 1.8	—	—
daf-16	Aspirin (1 mM)	32	14.5 $\pm$ 2.9	7	0.23
<i>Experiment 3</i>					
N2DRM	Control (vehicle)	35	15.1 $\pm$ 1.0	—	—
N2DRM	Aspirin (0.5 mM)	31	16.9 $\pm$ 2.4	12	< 0.01
N2DRM	Aspirin (1 mM)	33	17.8 $\pm$ 2.1	18	< 0.001
N2DRM	Salicylate (1 mM)	35	16.9 $\pm$ 1.0	12	< 0.01
<i>Sum of 2–3 Experiments:</i>		<i>Total N</i>	<i>Mean Increase</i>	<i>Composite p</i>	
N2DRM	Aspirin (0.5 mM)	166	23.0%	10 <sup>-8</sup>	
N2DRM	Aspirin (1 mM)	158	20.9%	10 <sup>-9</sup>	
N2DRM	Salicylate (1 mM)	164	13.9%	10 <sup>-6</sup>	
daf-16	Aspirin (1 mM)	66	3.4%	N.S.	

Mean adult lifespans are expressed in days after the L4/adult molt. Total lifespans are approximately 2.5 days longer (the developmental interval from egg-lay to adult) at 20°C. The mean increase (last 4 rows) has been weighted by the number of worms per experiment.

\*Significance of survival differences was ascertained by Gehans-Wilcoxon log-rank test, comparing treatment groups to untreated controls. The composite *p* value combining multiple independent experiments is the product of *p* values observed in the individual experiments.

implies that signaling through DAF-16/FOXO mediates most or all of aspirin's beneficial effects.

Both aspirin and salicylate slowed the age-dependent declines in motility and pharyngeal pumping, two of the most reliable physiological biomarkers of *C. elegans* aging (19, 24). Protein aggregation, due to oxidation, misfolding, and unstructured/nonpolar interactions between polyglutamine tracts, underlies aging-related neurodegenerative disorders and possibly many other age-dependent traits (30). Drugs that interfere with protein aggregation protect protein homeostasis, and can extend lifespan (1), and clinical trials indicate a substantial protective effect of long-term aspirin treatment against Alzheimer's dementia, which involves at least two types of protein aggregates (44). Using a nematode model of polyglutamine aggregation, we were able to demonstrate that aspirin indeed reduces aggregate size by >2-fold and the number of aggregates by >30% (Fig. 5). In view of the known association of protein aggregation with both oxidation and

aging (29), it is likely that the ability of aspirin to lower ROS levels and induce antioxidant defenses, demonstrated in the present study, is linked to inhibition of protein aggregation and extension of lifespan of the nematode.

The observation of *C. elegans* life-span extension by aspirin is consistent with the more modest increase in longevity observed for aspirin-treated male mice (43). In human subjects, aspirin ameliorated clinical parameters associated with type-2 diabetes (20), and moderately reduced hypertension (26). At therapeutic concentrations, aspirin is a potent anti-angiogenic agent and inhibits atherosclerotic plaque formation (23). Moreover, aspirin prevents restenosis after vascular injury, reduces the frequency of recurrent myocardial infarctions and strokes, and reduces or delays the development and progression of several cancers in humans (39). One plausible explanation for the remarkable versatility of aspirin in conferring protection against such a wide assortment of age-dependent diseases is that aspirin opposes insulin/IGF signaling, a pathway that contributes to aging and age-associated diseases (22), and/or opposes other pathways that converge on the DAF-16/FOXO transcription factor (49). The 21%–23% mean life extension observed in aspirin-treated nematodes, dependent on functional DAF-16, constitutes the strongest evidence to date for such a general, anti-aging effect. Since salicylate conferred benefits similar to those of aspirin, but by most measures was less effective, it appears that the beneficial effects of aspirin described here are mediated largely, but not entirely, by the salicylate moiety.

"Oxidative damage" theories of aging attribute its ravages to cell damage inflicted by ROS. Such theories have fallen into disfavor of late, due to the inability to forestall aging with genetic or drug interventions that demonstrably reduce markers of oxidative damage (12, 36, 46). Although cogently argued, these critiques challenge a rather narrow and outdated version of such theories. They thus fail to take into account the remarkable redundancy and compensatory ability of antioxidant defenses (4), and the positive role of oxygen free radicals in inflammatory signaling. Thus, an intervention that defeats any one antioxidant effector could induce other overlapping defenses, or might thwart inflammatory reactions that are critical for defense against pathogens (50).

## Materials and Methods

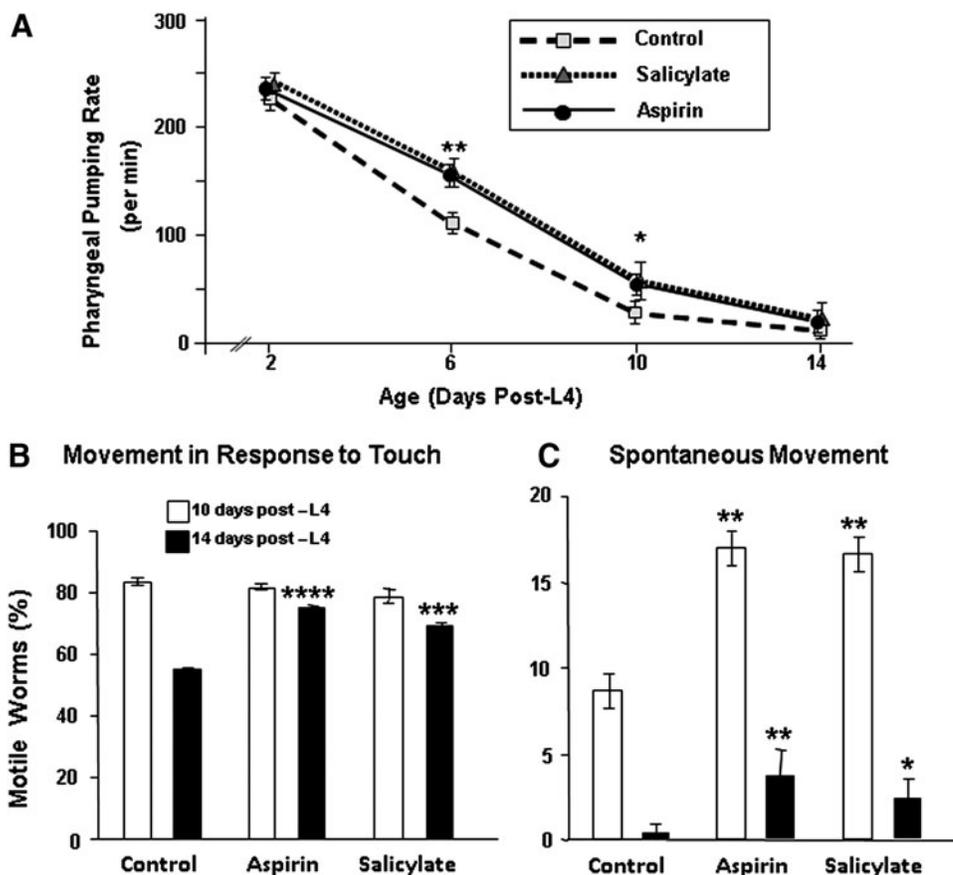
### Strains

Nematode strains, supplied by the Caenorhabditis Genetics Center (CGC, Minneapolis), or derived in our laboratory from CGC strains, were maintained at 20°C on 0.6% peptone NGM-agar plates seeded with *E. coli* strain OP50, as described earlier (2). Strains employed in these studies were Bristol-N2 (wild-type) and CF1038 bearing a *daf-16(mu86)* allele in which most of the exons encoding DAF-16 are deleted.

### Determination of lifespan

Nematodes, grown on NGM-agar plates containing 0.6% peptone, were harvested, and eggs were isolated by alkaline hypochlorite with 0.5 N NaOH, 1.05% hypochlorite; 5 min at 20°C (2). The recovered eggs were rinsed in S Buffer and placed on fresh agar plates seeded with *E. coli* strain OP50. Survival cultures were established on 60-mm agar plates; just after the L4/adult molt, 50 adults were transferred to 60-mm

**FIG. 4.** Age-dependent declines in activity are delayed by aspirin or salicylate, each at 0.5 mM. (A) Pharyngeal pumping rate declines with adult age. Significance of differences by 2-tailed *t* test (each  $N \geq 20$ ): \* $p < 0.01$  for aspirin vs. control at 10 days post-L4; \*\* $p < 0.001$  for either treatment vs. control at 6 days post-L4. (B) Movement in response to a touch stimulus drops ~30% between 10 and 14 days after the L4/adult molt (control bars), but this is largely blocked by aspirin or salicylate (each at 0.5 mM). (C) The spontaneous movement frequency at 10 days of adult age is doubled by either aspirin or salicylate; the very low level at 14 days is increased by an even larger factor. Significance by Chi-squared test, comparing treated to untreated control worms for each trait (B, C:  $N = 300$  per group): \* $p < 0.05$ ; \*\* $p < 0.005$ ; \*\*\* $p < 0.0005$ ; \*\*\*\* $p < 10^{-6}$ . Ages are given as days after the L4/adult molt.



dishes containing aspirin, salicylic acid, or ethanol (solvent control, 5% [w/v] final concentration) at the indicated concentrations. Worms were maintained at  $20 \pm 0.5^\circ\text{C}$  and live worms counted during daily transfer to fresh dishes; worms not moving, either spontaneously or in response to touch, were scored as dead. Worms were maintained until death.

#### Hydrogen peroxide stress response

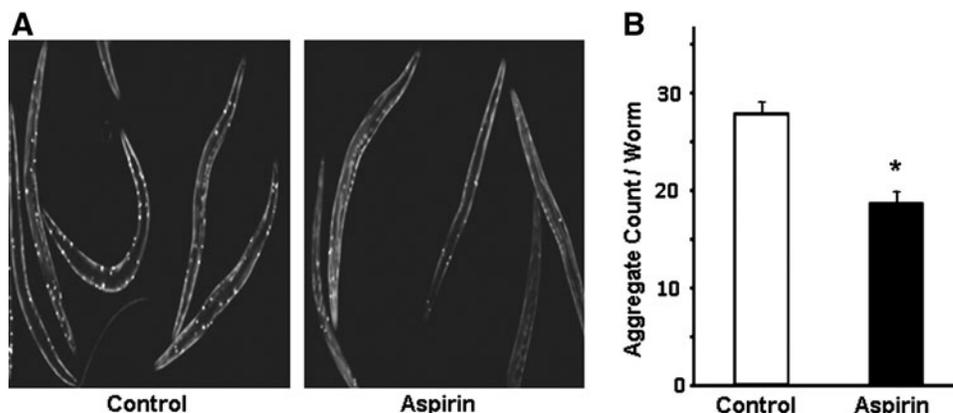
Adult worms (*N2* or *daf-16*) were synchronized by alkaline hypochlorite lysis, and then the eggs were rinsed and transferred to fresh NGM-agar plates. On reaching the L4 stage, worms were placed on fresh NGM-agar plates seeded with *E. coli* strain OP50 and overlaid with 1 mM aspirin in ethanol,

or ethanol alone (control). After 48 h, they were transferred to 24-well plates (20–25 worms per well) containing S Medium (S Buffer plus 0.5% cholesterol) and 5 mM hydrogen peroxide (Sigma) at  $20^\circ\text{C}$ , as previously described (2, 3). Survival was scored at 1-hour intervals, as above.

#### DCF assay for endogenous hydrogen peroxide

Endogenous ROS levels were indirectly quantified using dichlorodihydrofluorescein diacetate,  $\text{H}_2\text{DCF-DA}$  (Invitrogen), which is deacetylated and retained after uptake into cells. In turn,  $\text{H}_2\text{DCF}$  reacts with endogenous ROS to generate the fluorescent dye, dichlorofluorescein (DCF). Live *C. elegans* nematodes were treated at day 4 post-hatch (1.5 days

**FIG. 5.** Protein aggregates in *C. elegans* strain AM141. Aggregates in worms ~4 days after the L4/adult molt were reduced by life-long exposure to 1 mM aspirin. (A) Typical worms in each group; (B) Data summarized for  $\geq 30$  worms in each group. \* $p < 0.0001$  vs. control.



after the L4/adult molt) with 10  $\mu$ M H<sub>2</sub>DCF-DA for 30 min. Fluorescence intensity was imaged using an epifluorescence microscope (Olympus BX151) with 488-nm incident light, recording light emission at 520  $\pm$  20 nm with a QICAM camera. The intensity and duration of incident-light exposure was kept constant, including the extent of prior light exposure which might introduce photobleaching artefacts. Fluorescence intensity, recorded through a narrow-band-pass emission filter, was quantified from digital images using *imageJ* software.

#### Fluorescence measurement of GFP-reporter strains

A confocal epifluorescence microscope (Olympus model BX151) was used to image strains carrying P<sub>gst-10</sub>::GFP or P<sub>gst-4</sub>::GFP transgenes, with 390-nm excitation light, recording green epifluorescence at 510  $\pm$  20 nm. For each reporter strain, the same image-capture settings were utilized for treated and untreated control worms, using a QICAM. Autofluorescence was measured for non-GFP (background-control) worms of the same age, and subtracted from each image. Total GFP fluorescence per worm was analyzed as above, using *imageJ* software.

#### Locomotion, pharyngeal pumping, and protein aggregate measurement

Locomotory movement, either spontaneous or in response to touch, was measured for synchronized wild-type N2 cultures as described previously (16, 19). To assess protein aggregation, AM141 worms expressing a muscle-specific fusion protein, polyglutamine (Q40)::YFP, were synchronized and grown at 20°C as above. From the L4/adult interface onward, treated worms were maintained in the presence of aspirin, salicylate, or the ethanol solvent used to introduce those drugs to plates. Epifluorescence images were captured as above. Each worm was analyzed using *imageJ* software, and aggregates were counted in at least 30 animals per group.

#### Statistical analysis

Survival and stress test experiments were repeated at least three times. Median survival (either lifespan or survival of peroxide stress) was calculated from the fitted Gompertz function at 50% survival, using NCSS software (Number Cruncher Statistical Systems, Kaysville, UT). Significance of survival differences was determined by the Gehans-Wilcoxon log-rank test, a nonparametric measure that assesses differences in entire survival curves. For comparisons of proportions (e.g., for motility assays), the chi-squared test was utilized. Other comparisons between groups involved Student's *t* test for larger samples ( $N \geq 10$ ), or the Behrens-Fisher version of the *t* test appropriate to small samples in which equality of variances could not be established. *P* values  $\leq 0.05$  are reported as nominally significant in two-group comparisons, but sufficient information is provided for stringent adjustments in threshold to be made for multiple endpoint comparisons.

#### Acknowledgments

This study was supported in part by funds from the Department of Veterans Affairs, Veterans Health Administra-

tion, Office of Research and Development, Biomedical Laboratory Research and Development, Washington, DC. The contents of this article do not represent the views of the Department of Veterans Affairs or the United States Government.

#### Author Disclosure Statement

None of the authors declare any conflict of financial interest.

#### References

- Alavez S, Vantipalli MC, Zucker DJ, Klang IM, and Lithgow GJ. Amyloid-binding compounds maintain protein homeostasis during ageing and extend lifespan. *Nature* 472: 226–229, 2011.
- Ayyadevara S, Alla R, Thaden JJ, and Shmookler Reis RJ. Remarkable longevity and stress resistance of nematode PI3K-null mutants. *Aging Cell* 7: 13–22, 2008.
- Ayyadevara S, Dandapat A, Singh SP, Siegel ER, Shmookler Reis RJ, Zimniak L, and Zimniak P. Life span and stress resistance of *Caenorhabditis elegans* are differentially affected by glutathione transferases metabolizing 4-hydroxynon-2-enal. *Mech Ageing Dev* 128: 196–205, 2007.
- Back P, Matthijssens F, Vlaeminck C, Braeckman BP, and Vanfleteren JR. Effects of *sod* gene overexpression and deletion mutation on the expression profiles of reporter genes of major detoxification pathways in *Caenorhabditis elegans*. *Exp Gerontol* 45: 603–610, 2010.
- Balaban RS, Nemoto S, and Finkel T. Mitochondria, oxidants, and aging. *Cell* 120: 483–495, 2005.
- Brignull HR, Morley JF, Garcia SM, and Morimoto RI. Modeling polyglutamine pathogenesis in *C. elegans*. *Methods Enzymol* 412: 256–282, 2006.
- Bulckaen H, Prevost G, Boulanger E, Robitaille G, Roquet V, Gaxatte C, Garcon G, Corman B, Gosset P, Shirali P, Creusy C, and Puisieux F. Low-dose aspirin prevents age-related endothelial dysfunction in a mouse model of physiological aging. *Am J Physiol Heart Circ Physiol* 294: H1562–H1570, 2008.
- Campisi J and Yaswen P. Aging and cancer cell biology, 2009. *Aging Cell* 8: 221–225, 2009.
- Chavez V, Mohri-Shiomi A, Maadani A, Vega LA, and Garsin DA. Oxidative stress enzymes are required for DAF-16-mediated immunity due to generation of reactive oxygen species by *Caenorhabditis elegans*. *Genetics* 176: 1567–1577, 2007.
- Chen X, Zhong Z, Xu Z, Chen L, and Wang Y. 2',7'-Dichlorodihydrofluorescein as a fluorescent probe for reactive oxygen species measurement: Forty years of application and controversy. *Free Radic Res* 44: 587–604, 2010.
- De Berardis G, Sacco M, Strippoli GF, Pellegrini F, Graziano G, Tognoni G, and Nicolucci A. Aspirin for primary prevention of cardiovascular events in people with diabetes: Meta-analysis of randomised controlled trials. *BMJ* 339: b4531, 2009.
- Doonan R, McElwee JJ, Matthijssens F, Walker GA, Houthoofd K, Back P, Matscheski A, Vanfleteren JR, and Gems D. Against the oxidative damage theory of aging: Superoxide dismutases protect against oxidative stress but have little or no effect on life span in *Caenorhabditis elegans*. *Genes Dev* 22: 3236–3241, 2008.
- Franceschi C, Olivieri F, Marchegiani F, Cardelli M, Cavallone L, Capri M, Salvioli S, Valensin S, De Benedictis G, Di Iorio A, Caruso C, Paolisso G, and Monti D. Genes involved in immune response/inflammation, IGF1/insulin pathway and response to oxidative stress play a major role

- in the genetics of human longevity: The lesson of centenarians. *Mech Ageing Dev* 126: 351–361, 2005.
14. Guarente L and Kenyon C. Genetic pathways that regulate ageing in model organisms. *Nature* 408: 255–262, 2000.
  15. Harman D. Aging and oxidative stress. *J Int Fed Clin Chem* 10: 24–27, 1998.
  16. Herndon LA, Schmeissner PJ, Dudaronek JM, Brown PA, Listner KM, Sakano Y, Paupard MC, Hall DH, and Driscoll M. Stochastic and genetic factors influence tissue-specific decline in ageing *C. elegans*. *Nature* 419: 808–814, 2002.
  17. Holzenberger M, Dupont J, Ducos B, Leneuve P, Geloën A, Even PC, Cervera P, and Le Bouc Y. IGF-1 receptor regulates lifespan and resistance to oxidative stress in mice. *Nature* 421: 182–187, 2003.
  18. Honda Y and Honda S. The *daf-2* gene network for longevity regulates oxidative stress resistance and Mn-superoxide dismutase gene expression in *Caenorhabditis elegans*. *FASEB J* 13: 1385–1393, 1999.
  19. Huang C, Xiong C, and Kornfeld K. Measurements of age-related changes of physiological processes that predict lifespan of *Caenorhabditis elegans*. *Proc Natl Acad Sci USA* 101: 8084–8089, 2004.
  20. Hundal RS, Petersen KF, Mayerson AB, Randhawa PS, Inzucchi S, Shoelson SE, and Shulman GI. Mechanism by which high-dose aspirin improves glucose metabolism in type 2 diabetes. *J Clin Invest* 109: 1321–1326, 2002.
  21. Johnson TE, Cypser J, de CE, de CS, Henderson S, Murakami S, Rikke B, Tedesco P, and Link C. Gerontogenes mediate health and longevity in nematodes through increasing resistance to environmental toxins and stressors. *Exp Gerontol* 35: 687–694, 2000.
  22. Kenyon C. A conserved regulatory system for aging. *Cell* 105: 165–168, 2001.
  23. Khaidakov M, Szewedo J, Mitra S, Ayyadevara S, Dobretsov M, Lu J, and Mehta JL. Antiangiogenic and antimitotic effects of aspirin in hypoxia-reoxygenation modulation of the LOX-1-NADPH oxidase axis as a potential mechanism. *J Cardiovasc Pharmacol* 56: 635–641, 2010.
  24. Lakowski B and Hekimi S. The genetics of caloric restriction in *Caenorhabditis elegans*. *Proc Natl Acad Sci USA* 95: 13091–13096, 1998.
  25. Lin K, Hsin H, Libina N, and Kenyon C. Regulation of the *Caenorhabditis elegans* longevity protein DAF-16 by insulin/IGF-1 and germline signaling. *Nature Genetics* 28: 139–145, 2001.
  26. Magen E, Viskoper JR, Mishal J, Priluk R, London D, and Yosefy C. Effects of low-dose aspirin on blood pressure and endothelial function of treated hypertensive hypercholesterolaemic subjects. *J Hum Hypertens* 19: 667–673, 2005.
  27. McElwee JJ, Schuster E, Blanc E, Thomas JH, and Gems D. Shared transcriptional signature in *Caenorhabditis elegans* Dauer larvae and long-lived *daf-2* mutants implicates detoxification system in longevity assurance. *J Biol Chem* 279: 44533–44543, 2004.
  28. Mehta JL, Chen J, Yu F, and Li DY. Aspirin inhibits ox-LDL-mediated LOX-1 expression and metalloproteinase-1 in human coronary endothelial cells. *Cardiovasc Res* 64: 243–249, 2004.
  29. Morimoto RI. Stress, aging, and neurodegenerative disease. *N Engl J Med* 355: 2254–2255, 2006.
  30. Morley JF, Brignull HR, Weyers JJ, and Morimoto RI. The threshold for polyglutamine-expansion protein aggregation and cellular toxicity is dynamic and influenced by aging in *Caenorhabditis elegans*. *Proc Natl Acad Sci USA* 99: 10417–10422, 2002.
  31. Muller FL, Song W, Jang YC, Liu Y, Sabia M, Richardson A, and Van RH. Denervation-induced skeletal muscle atrophy is associated with increased mitochondrial ROS production. *Am J Physiol Regul Integr Comp Physiol* 293: R1159–R1168, 2007.
  32. Murphy CT, McCarroll SA, Bargmann CI, Fraser A, Kamath RS, Ahringer J, Li H, and Kenyon C. Genes that act downstream of DAF-16 to influence the lifespan of *Caenorhabditis elegans*. *Nature* 424: 277–283, 2003.
  33. Nguyen T, Sherratt PJ, and Pickett CB. Regulatory mechanisms controlling gene expression mediated by the antioxidant response element. *Annu Rev Pharmacol Toxicol* 43: 233–260, 2003.
  34. Okuyama T, Inoue H, Ookuma S, Satoh T, Kano K, Honjoh S, Hisamoto N, Matsumoto K, and Nishida E. The ERK-MAPK pathway regulates longevity through SKN-1 and insulin-like signaling in *Caenorhabditis elegans*. *J Biol Chem* 285: 30274–30281, 2010.
  35. Ong G, Davis TM, and Davis WA. Aspirin is associated with reduced cardiovascular and all-cause mortality in type 2 diabetes in a primary prevention setting: The Fremantle Diabetes study. *Diabetes Care* 33: 317–321, 2010.
  36. Perez VI, Bokov A, Van RH, Mele J, Ran Q, Ikeno Y, and Richardson A. Is the oxidative stress theory of aging dead? *Biochim Biophys Acta* 1790: 1005–1014, 2009.
  37. Redondo S, Santos-Gallego CG, Ganado P, Garcia M, Rico L, Del Rio M, and Tejerina T. Acetylsalicylic acid inhibits cell proliferation by involving transforming growth factor-beta. *Circulation* 107: 626–629, 2003.
  38. Rozing MP, Westendorp RG, Frolich M, de Craen AJ, Beekman M, Heijmans BT, Mooijaart SP, Blauw GJ, Slagboom PE, and van Heemst D. Human insulin/IGF-1 and familial longevity at middle age. *Ageing (Albany NY)* 1: 714–722, 2009.
  39. Schreinemachers DM and Everson RB. Aspirin use and lung, colon, and breast cancer incidence in a prospective study. *Epidemiology* 5: 138–146, 1994.
  40. Shmookler Reis RJ, Kang P, and Ayyadevara S. Quantitative trait loci define genes and pathways underlying genetic variation in longevity. *Exp Gerontol* 41: 1046–1054, 2006.
  41. Shmookler Reis RJ, Xu L, Lee H, Chae M, Thaden JJ, Barill P, Tazearslan C, Siegel E, Alla R, Zimniak P, and Ayyadevara S. Modulation of lipid biosynthesis contributes to stress resistance and longevity of *C. elegans* mutants. *Ageing (Albany NY)* 3: 125–147, 2011.
  42. Sohal RS, Mockett RJ, and Orr WC. Mechanisms of aging: An appraisal of the oxidative stress hypothesis. *Free Radic Biol Med* 33: 575–586, 2002.
  43. Strong R, Miller RA, Astle CM, Floyd RA, Flurkey K, Hensley KL, Javors MA, Leeuwenburgh C, Nelson JF, Ongini E, Nadon NL, Warner HR, and Harrison DE. Nordihydroguaiaretic acid and aspirin increase lifespan of genetically heterogeneous male mice. *Ageing Cell* 7: 641–650, 2008.
  44. Thomas T, Nadackal TG, and Thomas K. Aspirin and non-steroidal anti-inflammatory drugs inhibit amyloid-beta aggregation. *Neuroreport* 12: 3263–3267, 2001.
  45. Valko M, Leibfritz D, Moncol J, Cronin MT, Mazur M, and Telser J. Free radicals and antioxidants in normal physiological functions and human disease. *Int J Biochem Cell Biol* 39: 44–84, 2007.
  46. Van Raamsdonk JM and Hekimi S. Reactive oxygen species and aging in *Caenorhabditis elegans*: Causal or casual relationship? *Antioxid Redox Signal* 13: 1911–1953, 2010.
  47. Vasto S, Carruba G, Lio D, Colonna-Romano G, Di Bona D, Candore G, and Caruso C. Inflammation, ageing and cancer. *Mech Ageing Dev* 130: 40–45, 2009.
  48. Vissinger H, Husted SE, Kristensen SD, and Nielsen HK. Platelet-derived growth factor release and antiplatelet treatment with low-dose acetylsalicylic acid. *Angiology* 44: 633–638, 1993.

49. Yen K, Narasimhan SD, and Tissenbaum HA. DAF-16/Forkhead box O transcription factor: Many paths to a single Fork (head) in the road. *Antioxid Redox Signal* 14: 623–634, 2011.
50. Zimniak P. Detoxification reactions: Relevance to aging. *Ageing Res Rev* 7: 281–300, 2008.

Date of first submission to ARS Central, August 31, 2011; date of final revised submission, August 5, 2012; date of acceptance, August 6, 2012.

Address correspondence to:

Dr. Srinivas Ayyadevara  
VA Medical Center and UAMS  
4300 West 7<sup>th</sup> Street, Research LR/151  
Little Rock, AR 72205

E-mail: AyyadevaraSrinivas@uams.edu

or

Dr. J.L. Mehta  
Univ. of Arkansas for Medical Sciences (UAMS)  
4301 W Markham, Slot 532  
Little Rock, AR 72205

E-mail: mehtajl@uams.edu

#### Abbreviations Used

*C. elegans* = *Caenorhabditis elegans*  
CTL = catalase  
DAF-16 = Dauer formation protein 16  
FOXO = forkhead box protein O  
GSTs = glutathione S-transferases  
H<sub>2</sub>DCF-DA = dichlorodihydrofluorescein diacetate  
IGF-1 = insulin-like growth factor-1  
PDGF = platelet-derived growth factor  
ROS = reactive oxygen species  
SOD = superoxide dismutase  
TGF- $\beta$  = transforming growth factor- $\beta$   
UGT = UDP glucuronosyltransferase